

**Abbreviations**

8-dG: 8-hydroxydesoxyguanosine; AFC: EFSA Panel on food additives; flavourings: processing aids and materials in contact with food; ALT: alanine aminotransferase; APFO: ammonium perfluorooctanoate; AST: aspartate aminotransferase; BfR: Bundesinstitut für Risikobewertung (Federal Institute for Risk Assessment); BMI: body mass index; BW: body weight; CAR: constitutive androstane receptor; COT: Committee on Toxicity of Chemicals in Food: Consumer Products and the Environment; ECF: electrochemical fluorination; EFSA: European Food Safety Authority; FDA: US Food and Drug Administration; FOSA: perfluorooctyl sulfonamide; FOSE: perfluorooctanesulfonamide ethanol; FT<sub>4</sub>: free thyroxine; FTOH: fluorotelomer alcohol; GD: gestational day; GJIC: gap junction intercellular communication; HDL: high density lipoprotein; LADD: lifetime average daily dose; LC<sub>50</sub>: lethal concentration with 50% lethality; LD<sub>50</sub>: lethal dose with 50% lethality; L-FABP: liver type fatty acid binding protein; LH: luteinizing hormone; LOAEL: lowest observed adverse effect level; L-PFOA: linear PFOA; L-PFOS linear PFOS; MTE: more typical exposure; N (or n): number (of samples); n.a.: not analyzed; n.d.: not detected; n.r.: not reported; N-EtFOSA: N-ethylperfluorooctyl sulfonamide; N-EtFOSE: N-ethylperfluorooctyl sulfonamide ethanol; NHANES: National Health and Nutrition Examination Survey; NOAEL: no observed adverse effect level; OAT: organic anion transporter; OECD: Organisation for Economic Co-operation and Development; PCB: polychlorinated biphenyls; PFBA: perfluorobutanoic acid; PFBS: perfluorobutanesulfonic acid; PFCs: perfluorinated compounds/polyfluorinated compounds; PFCA: perfluorinated carboxylic acids; PFDA: perfluorodecanoic acid; PFDDA: perfluorododecanoic acid; PFHpA: perfluoroheptanoic acid; PFHxA: perfluorohexanoic acid; PFHxS: perfluorohexanesulfonic acid; PFNA: perfluorononanoic acid; PFO: deprotonated PFOA; PFOA: perfluorooctanoic acid; PFOS: perfluorooctanesulfonic acid; PFPA: perfluorinated phosphonic acid; PFPeA: perfluoropentanoic acid; PFTDA: perfluorotetradecanoic acid; PFUnA: perfluoroundecanoic acid; POP: persistent organic pollutants; POSF: perfluorooctane sulfonyl fluoride; PPAR: peroxisome proliferator activated receptor; PTFE: polytetrafluoroethylene; PXR: pregnane x receptor; RME: reasonable maximum exposure; ROS: reactive oxygen species; SMR: standard mortality ratio; SOD: superoxide dismutase; T<sub>3</sub>: tri-iodo thyronine; T<sub>4</sub>: thyroxine; T-AOC: total antioxidant capacity; TDI: tolerable daily intake; TSH: thyroid stimulating hormone; UBA: Umweltbundesamt (German Environmental Protection Agency); US EPA: United States Environmental Protection Agency.

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**Authors' contributions**

TS had the idea to write such a review and drafted the manuscript. DM evaluated the original literature and participated in drafting the manuscript. HB structured this complex review in chapters and subchapters and participated in drafting the manuscript. All authors read and approved the final manuscript.

**Competing interests**

The authors declare that they have no competing interests.

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## Deriving environmental quality standards for perfluorooctanoic acid (PFOA) and related short chain perfluorinated alkyl acids

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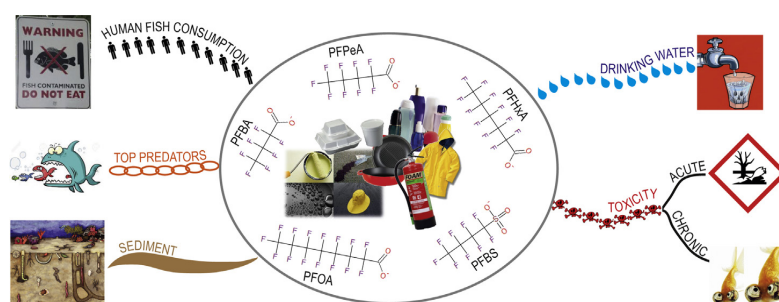
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### HIGHLIGHTS

- Environmental Quality Standards (EQS) for selected perfluoroalkylacids are derived.
- For PFBA, PFPeA, PFHxA and PFBS the QS for drinking water are adopted as EQS.
- For PFOA the QS based on secondary poisoning of predators is adopted as EQS.
- 7, 3, 1, 3 and 0.1  $\mu\text{gL}^{-1}$  for PFBA, PFPeA, PFHxA, PFBS and PFOA are set as EQS.

### GRAPHICAL ABSTRACT



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### ABSTRACT

The evidence that in Northern Italy significant sources of perfluoroalkylacids (PFAA) are present induced the Italian government to establish a Working Group on Environmental Quality Standard (EQS) for PFAA in order to include some of them in the list of national specific pollutants for surface water monitoring according to the Water Framework Directive (2000/60/EC). The list of substances included perfluorooctanoate (PFOA) and related short chain PFAA such as perfluorobutanoate (PFBA), perfluoropentanoate (PFPeA), perfluorohexanoate (PFHxA) and perfluorobutanesulfonate (PFBS), which is a substitute of perfluorooctanesulfonate. For each of them a dossier collects available data on regulation, physico-chemical properties, emission and sources, occurrence, acute and chronic toxicity on aquatic species and mammals, including humans.

Quality standards (QS) were derived for the different protection objectives (pelagic and benthic communities, predators by secondary poisoning, human health via consumption of fishery products and water) according to the European guideline. The lowest QS is finally chosen as the relevant EQS. For PFOA a QS for biota was derived for protection from secondary poisoning and the corresponding QS

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for water was back-calculated, obtaining a freshwater EQS of  $0.1 \mu\text{g L}^{-1}$ . For PFBA, PFPeA, PFHxA and PFBS threshold limits proposed for drinking waters were adopted as EQS.

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## 1. Introduction

Because of their peculiar physical and chemical characteristics, perfluoroalkyl substance (PFAS) found wide application in several industrial processes and products such as surface treatment of textiles and paper, building paints, cosmetics, insecticide formulations, firefighting foams, and the production of fluoropolymer. Their characteristics such as resistance to hydrolysis, photolysis, microbial degradation make these substances highly persistent and widespread in all environmental compartments, especially water. PFAS include thousands of chemicals but environmental studies have mainly concentrated on perfluoroalkyl acids (PFAA) such as perfluoroalkylsulfonic acids (PFSA) and perfluoroalkylcarboxylic acids (PFCA). PFAA are low molecular weight surfactants in which all carbons are bonded to fluorine atoms, consisting of homologous series of molecules that differ in carbon chain length. The two PFAS most commonly used and found in the environment are perfluorooctanesulfonate (PFOS) and perfluorooctanoate (PFOA). They are widely employed in different industrial processes and their persistence in the environment associate with bioaccumulation in the trophic chain raising concern about the risks for consumers, including humans. Therefore the European Commission recently included PFOS in the list of priority substances, setting an Environmental Quality Standard (EQS) of  $0.65 \text{ ng L}^{-1}$  for freshwater and  $9.1 \text{ ng g}^{-1}$  for biota (Directive 2013/39/EC).

The hazard profile of PFOA is well known: PFOA is a persistent, bioaccumulative, and toxic (PBT) substance, which may cause severe and irreversible adverse effects on the environment and human health. PFOA has a harmonised classification in Annex VI of European Regulation (EC) No 1272/2008 on classification, labelling and packaging of substances and mixtures (CLP) as Carc. 2, Repr. 1B and STOT RE 1 (liver). Due to its PBT and CMR properties, PFOA and its ammonium salt (APFO) have been identified as substances of very high concern (SVHC) under REACH by unanimous agreement between EU Member States in July 2013.

At the moment for PFOA, no limits have been established in the aquatic environment but only a provisional threshold for drinking waters was proposed by US Environmental Protection Agency (EPA) of  $0.4 \mu\text{g L}^{-1}$  (EPA, 2009). Shorter chain (<C7) homologues are currently tested and used as substitutes for PFOS and PFOA in industrial processes.

The evidence that in Northern Italy significant sources of PFAAs are present and surface and ground-waters are significantly impacted [2,3] induced the Italian government to establish a Working Group on Environmental Quality Standard (EQS) for PFAAs in order to include some of them in the list of national specific pollutants for surface water monitoring and derive their EQSs for water quality assessment in the context of the Water Framework Directive (2000/60/EC). EQSs should protect freshwater and marine ecosystems from possible short and long-term adverse effects of chemicals as well as human health via drinking water or ingestion of food originating from aquatic environments.

The PFFAs were chosen by considering the statistical distribution of concentrations and frequency of detection in the Italian surface and ground waters [3,4]. The list included the following substances: PFOA and related short chain (number of carbons <7) perfluorocarboxylate namely perfluorobutanoate (PFBA), perfluoropentanoate (PFPeA) and perfluorohexanoate (PFHxA).

Perfluorobutanesulfonate (PFBS), which is a common substitute of PFOS, was considered too.

For each of them a dossier has been prepared which collects available data on regulation, physico-chemical properties, emission and sources, occurrence, acute and chronic toxicity on aquatic species and mammals, including humans.

This paper aims to present how EQSs for selected perfluoroalkylacids PFOA, PFBA, PFPeA, PFHxA and PFBS have been derived according to the European guidance document for derivation of EQSs under the Water Framework Directive [1]. By using the data in the dossiers, Quality Standards (QSs) were derived for different protection objectives based on: direct ecotoxicity to pelagic aquatic organisms; secondary poisoning of predators, human consumption of fishery products, human consumption of drinking water and to protect benthic (sediment-dwelling) species. Dossiers and proposed QSs were subject to a peer-reviewing process involving national experts of the Ministry of the Environment and international experts from other European Member States. After the reviewing process, the most protective QSs have been chosen as the national EQSs for the selected perfluoroalkylacids and adopted in the Italian legislation (Italian Legislative Decree 172/2015).

## 2. Methodology

EQSs were derived for PFOA and selected short chain perfluoroalkyl compounds PFBA, PFPeA, PFHxA and PFBS (referred to as SC-PFAA in the following text) in accordance with the European technical guidance document for derivation of environmental quality standards under the WFD (TGD-EQS) [1].

For each compound a dossier was compiled (Supplementary materials, SM1–SM5) according to the TGD-EQS format. This format requires regulatory information, physico-chemical data, occurrence in the different aquatic matrices, ecotoxicological data and toxicity data on mammals.

Toxicity data were collected from the scientific literature and from national and international databases. All available studies are summarised in the tables provided as Supplementary Materials (Tables SM7–SM9, with references in SM10). No toxicological data about invertebrates exposed to contaminated sediment have been found for the compounds under investigation.

The whole derived data set was properly assessed for reliability and relevance according to Klimisch et al. [5]. On this basis a selection and aggregation of the data were performed.

In order to cover both short- and long-term effects resulting from exposure, two water column EQSs is required: a short-term standard, referred to as a Maximum Allowable Concentration EQS (MAC-EQS) and a long-term standard, expressed as Annual Average Concentration EQS (AA-EQS). The MAC-EQS is based on information on direct ecotoxicological effects on aquatic organisms only. As AA-EQS is intended to protect the ecosystem, humans and predatory birds and mammals from long-term risks, it is defined as the lowest value among the derived QSs, calculated for the different protection objectives.

When the data were sufficient, the QSs were derived by probabilistic approach adopting Species Sensitivity Distribution (SSD) modelling. Otherwise, a deterministic approach was used applying appropriate assessment actors (AF) to the lowest reliable toxicity data.

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**Glossary**

AA-EQS <sub>fw,eco</sub> or AA-EQS <sub>sw,eco</sub>	Annual Average Environmental Quality Standard in fresh or salt water [ $\mu\text{g L}^{-1}$ ]
AA-QS <sub>fw,eco</sub> or AA-QS <sub>sw,eco</sub>	QS based on direct ecotoxicity to pelagic organisms in fresh or salt water [ $\mu\text{g L}^{-1}$ ]
ADI	Acceptable Daily Intake [ $\text{mg kg}_{\text{bw}}^{-1} \text{d}^{-1}$ ]
AF	Assessment Factor
AF <sub>oral</sub>	Assessment Factor applied in extrapolation of QS <sub>biota,secpois</sub>
BAF	Bioaccumulation Factor [ $\text{L kg}^{-1}$ ]
BCF	Bioconcentration Factor [ $\text{L kg}^{-1}$ ]
BMDL10	The lower limit of the 95% confidence interval of the benchmark dose for a 10% increase in effects [ $\mu\text{g kg}_{\text{bw}}^{-1} \text{d}^{-1}$ ]
BMF1	Biomagnification Factor of a simple food web that consists of water –BCF→ aquatic organisms –BMF1→ fish → fish-eating predator
BMF2	Biomagnification Factor of a marine simple food web that consists of water –BCF→ aquatic organisms –BMF1→ fish –BMF2→ fish-eating predator → top predator
bw	Body weight [kg]
DFI	Daily Food Intake [ $\text{kg d}^{-1}$ ]
EQS	Environmental Quality Standard
HC5	Hazardous concentration for 5% of the species (based on the SSD)
L(E)C50	One-half (50%) of the maximal Lethal (Effective) Concentration
LOAEL	Lowest Observed Adverse Effect Levels [ $\text{mg kg}_{\text{bw}}^{-1} \text{d}^{-1}$ ]
MAC-QS <sub>fw,eco</sub> or MAC-QS <sub>sw,eco</sub>	Maximum Allowable Concentration in fresh or salt water (short-term QS based on acute toxicity data) [ $\mu\text{g L}^{-1}$ ]
NOAEL	No Observed Adverse Effect Level [ $\text{mg kg}_{\text{bw}}^{-1} \text{d}^{-1}$ ]
NOEC	No Observed Effect Concentration [ $\mu\text{g L}^{-1}$ or $\mu\text{g kg}^{-1}$ ]
PFAA	Perfluoroalkylacids
PFBA	Perfluorobutanoate
PFBS	Perfluorobutanesulfonate
PFHxA	Perfluorohexanoate
PFOA	Perfluorooctanoate
PFOS	Perfluorooctanesulfonate
PFPeA	Perfluoropentanoate
QC	Quality Criteria
QS	Quality Standard
QS <sub>biota,hh</sub>	QS based on human consumption of fishery products [ $\mu\text{g kg}^{-1}$ ]
QS <sub>biota,secpois,fw</sub> or QS <sub>biota,secpois,sw</sub>	QS based on secondary poisoning of predators for freshwater or saltwater compartment [ $\mu\text{g kg}^{-1}$ ]
QS <sub>dw,hh</sub>	QS for human consumption of drinking water [ $\mu\text{g L}^{-1}$ ]
QS <sub>fw,secpois</sub> or QS <sub>sw,secpois</sub>	QS based on secondary poisoning converted into equivalent freshwater or saltwater concentration [ $\mu\text{g L}^{-1}$ ]
QS <sub>sed,fw</sub> or QS <sub>sed,sw</sub>	Sediment quality standard based on direct ecotoxicity to benthic (sediment dwelling) organisms in fresh or salt water [ $\mu\text{g kg}^{-1}$ ]
SC-PFAA	Short chain (number of carbon <7) perfluoroalkyl acids (PFBA, PFPeA, PFHxA, PFBS)

SSD	Species Sensitivity Distribution model
TDI	Tolerable Daily Intake [ $\text{mg kg}_{\text{bw}}^{-1} \text{d}^{-1}$ ]
TGD-EQS	Technical Guidance for Deriving Environmental Quality Standards [1]
TL	Threshold Level
TOX <sub>oral</sub>	NOEC <sub>oral,bird</sub> or NOEC <sub>oral,mammals</sub> in $\text{kg kg}_{\text{food}(fw)}^{-1}$
WFD	Water Framework Directive (2000/60/EC)

The QSs for SC-PFAAs were determined using the deterministic approach. Only for PFOA the acute toxicity data were sufficient for statistical extrapolation of QS by implementing the SSD approach.

**3. Results and discussion****3.1. Toxicity data**

Among the compounds of interest the largest dataset has been collected for PFOA, which includes both acute and chronic toxicity tests on organisms exposed to water and toxicity test on mammals and birds (Tables 1 and 4, SM7, SM8 and SM9).

For the other short chain perfluoroalkyl acids, data set is much more limited. Acute toxicity data on aquatic organisms are available for all the SC-PFAA (Table 2, Table 3 and SM7) but chronic toxicity data to aquatic organisms are available only for PFBS (Table 4 and SM8). In the case of PFHxA the number of reliable data is insufficient to derive a consistent quality standard for pelagic organisms.

Few toxicity studies on mammalian diet and oral exposure are reported for PFBA, PFHxA and PFBS and only one mammalian toxicity study was found for PFPeA (SM9). For PFBA, PFPeA and PFHxA the number of studies is insufficient to derive a reliable QS<sub>biota,secpois</sub>. However indicative values of the biota standards for PFBA and PFHxA are derived based on the lowest NOAEL/LOAEL available.

Saltwater ecotoxicity data are scarce and insufficient to enable a statistical comparison of the freshwater and saltwater data and to define marine quality standards based only on saltwater toxicity values. The available data, however, do not point out a difference in sensitivity, therefore, according to TGD-EQS [1], the freshwater and saltwater data are combined and separate QSs for all selected PFAAs were derived for freshwater and saltwater environments using different assessment factors (TGD-EQS). This is consistent with the provisions of REACH for marine effects assessment where a larger AF is recommended to cover the increased uncertainty resulting from the larger diversity of marine ecosystems and the limited availability of effects data for marine life forms. No toxicological data of invertebrates exposed to contaminated sediment were available for any examined compound.

**3.2. Derivation of QS from direct toxicity in the water column****3.2.1. MAC-QS<sub>water,eco</sub> for PFOA**

Short term or acute toxicity tests are used to derive a Maximum Acceptable Concentration-QS (Table 1). Short term toxicity PFOA dataset contains reliable L(E)C50 values from 24 different species covering 8 taxonomic groups (including algae, crustacean, rotifers, insect, annelids, echinodermata, amphibian and fish). The range of taxonomic groups covered and the number of L(E)C50 values from different species fulfil the TGD-EQS requirements for the use of the SSD to derive the MAC-QS. Acute toxicity data have been aggregated to one toxicity value per species, unreliable tests [5] were excluded from elaboration and SSD generator V<sub>1-12</sub> [6] has been used to calculate the hazardous concentration for 5% of species (HC5). Details on data selection and SSD elaboration are reported in Supplemental Materials (Table SM6, with references in SM10). The estimated

**Table 1**  
PFOA acute aquatic toxicity (The full dataset is available in Table SM7, with references in SM10).

	Species	Exp time	Endpoint	LC50 (95% CI) mg L <sup>-1</sup>	EC50 (95% CI) mg L <sup>-1</sup>	References
Freshwater						
	Cyanobacteria					
	<i>Geitlerinema amphibium</i>	72 h	biomass <sup>a</sup>		247.8 ± 12.4 <sup>b</sup>	[45]
	<i>Anabaena</i> CPB4337	24 h	luminescence inhibition		19.81 (15.44–26.44)	[46]
Algae	<i>Pseudochirchneriella subcapitata</i>	72 h	growth rate and biomass		>100	[47]
		96 h			>100	
	<i>Pseudochirchneriella subcapitata</i>	72 h	growth rate and biomass		>400	[11]
		96 h			>400	
	<i>Pseudochirchneriella subcapitata</i>	72 h	growth inhibition (biomass)		96.2 (88.6–113.7)	[48]
	<i>Pseudochirchneriella subcapitata</i>	4.5 h	photosynthesis		746.40 (726.99–768.29)	[49]
	<i>Scenedesmus quadricanda</i>	96 h	not reported		269.63 (207.83–349.82)	[50]
	<i>Chlamydomonas reinhardtii</i>	96 h	growth inhibition		51.9 ± 1.0	[51]
	<i>Scenedesmus obliquus</i>	96 h	growth inhibition		44.0 ± 1.5	[51]
	<i>Chlorella vulgaris</i>	72 h	biomass <sup>a</sup>		974.82 ± 49.56 <sup>b</sup>	[45]
Invertebrata: Crustacean	<i>Daphnia magna</i>	48 h	immobilization		480	[11]
	<i>Daphnia magna</i>	48 h	immobilization		480	[47]
	<i>Daphnia magna</i>	24 h	immobilization		675.05 (559.62–790.50)	[52]
		48 h			476.52 (375.32–577.72)	
	<i>Daphnia magna</i>	24 h	immobilization		219.33 (209.01–229.25)	[53]
		48 h			211.07 (184.22–254.86)	
	<i>Chydorus sphaericus</i>	24 h	immobilization		175.96 (92.11–221.81)	
		48 h			116.48 (50.39–142.51)	
	<i>Moina macrocopa</i>	24 h	immobilization		348.76 (272.65–424.87)	[52]
		48 h			199.51 (153.89–245.13)	
	<i>Daphnia magna</i>	48 h	mortality	201.85 (134.68–302.50)		[50]
	<i>Daphnia magna</i>	24 h	immobilization		298 (278–321)	[54]
		48 h			181 (166–198)	
	<i>Neocaridina denticulate</i> (green neon shrimp)	24 h	mortality	>1000 712(663–764)		[54]
	48 h		546 (502–594)			
	72 h		454 (418–494)			
<i>Macrobrachium nipponense</i> (freshwater shrimp)	96 h	mortality	366.66 (253.09–531.18)		[50]	
	96 h					
Invertebrata: Rotifers						
	<i>Brachionus calyciflorus</i>	24 h	mortality	150		[55]



Table 1 (Continued)

	Species	Exp time	Endpoint	LC50 (95% CI) mg L <sup>-1</sup>	EC50 (95% CI) mg L <sup>-1</sup>	References
Invertebrata: Gastropoda	<i>Physa acuta</i>	24 h	mortality	856 (768–954)		[54]
		48 h		732 (688–779)		
		72 h		697 (661–735)		
		96 h		672 (635–711)		
	<i>Cipangopaludina cathayensis</i>	96 h	mortality	740.07 (597.66–916.41)	[50]	
Invertebrata: Platyhelminthes	<i>Dugesia japonica</i>	24 h	mortality	352 (331–374)		[54]
		48 h		345 (325–366)		
		72 h		343 (324–364)		
		96 h		337 (318–357)		
Invertebrata: Insecta	<i>Chironomus plumosus</i>	96 h	mortality	402.24 (323.83–499.63)		[50]
Invertebrata: Anellida	<i>Limnodrilus hoffmeisteri</i>	96 h	mortality	568.20 (476.33–677.80)		[50]
Vertebrata: Amphibia	<i>Bufo gargarizans</i>	96 h	mortality	114.74 (83.02–158.58)		[50]
Vertebrata: Fish	<i>Pseudorasbora parva</i>	96 h	mortality	365.02 (269.78–493.86)		[50]
		96 h		606.61 (460.93–798.32)		
	<i>Carassius auratus</i>	96 h	mortality	707		[11]
		96 h		800		
		144 hpf		430 (290–710)		
	<i>Danio rerio</i> (zebrafish embryo test)	96 hpf	mortality and malformation effects mortality and malformation effects <sup>c</sup>	>500	350 (290–430)	[57]
		120 hpf		>500	205.72(168.25–251.53) 113.05 (96.22–132.84)	
Marine water Bacteria	<i>Vibrio fischeri</i>	30 min	luminescence inhibition		570.19 ± 57.33	[58]
		15 min	luminescence inhibition		524 (505–538)	[48]
	<i>Photobacterium phosphoreum</i>	15 min	luminescence inhibition		14.65	[7]
Algae	<i>Skeletonema marinoi</i>	72 h	biomass <sup>a</sup>		367.52 ± 16.5 <sup>b</sup>	[45]
		72 h	growth inhibition		163.6 (131.7–203.2)	[59]
Invertebrata: Crustacea	<i>Siriella armata</i>	96 h	mortality	15.5 (13.0–18.6)		[59]
Invertebrata: Echinodermata	<i>Paracentrotus lividus</i>	48 h	growth inhibition		110.0 (99.2–121.9)	[59]
Vertebrata: Fish	<i>Psetta maxima</i>	144 h	abnormalities/mortality	11.9 (9.5–14.9)		[59]

Notes: hpf: hours post fertilization.

<sup>a</sup> biomass assessed by optical density.<sup>b</sup> standard deviation.<sup>c</sup> malformation of head and tail and effects on growth.

**Table 2**  
PFBA, PFPeA, PFHxA acute aquatic toxicity (The full dataset is available in Table SM7, with references in SM10).

	Species	Exp time	Endpoint	LC50 (95% CI) mg L <sup>-1</sup>	EC50 (95% CI) mg L <sup>-1</sup>	References
Freshwater						
PFBA						
Algae	<i>Pseudokirchneriella subcapitata</i>	4.5 h	photosynthesis		260.96 (213.03–319.55) <sup>a</sup>	[49]
Invertebrata: Crustacean	<i>Daphnia magna</i>	24 h	immobilization		184.27 (182.78–185.55) <sup>a</sup> >4260.6 <sup>b</sup>	[53]
		48 h			180.65 (179.15–182.35) <sup>a</sup> >4260.6 <sup>b</sup>	
	<i>Chydorus sphaericus</i>	24 h	immobilization		534.49 (513.19–555.37) <sup>a</sup> >4.26 <sup>b</sup>	[53]
		48 h			460.14 (440.97–479.10) <sup>a</sup> >4260.6 <sup>b</sup>	
Invertebrata: Rotifera	<i>Brachionus calyciflorus</i>	24 h	mortality	110 <sup>a</sup>		[8]
Vertebrata: Fish	<i>Danio rerio</i>	144 hpf	mortality	>3000 <sup>a</sup>		[56]
			mortality and malformation effects		>2200 (1200–22,000) <sup>a,d</sup>	
	<i>Danio rerio</i>	96 hpf	mortality and malformation effects <sup>e</sup>	>3000 <sup>b</sup>		[57]
		120 hpf		>3000 <sup>b</sup>		
PFPeA						
Algae	<i>Pseudokirchneriella subcapitata</i>	72 h	biomass		81.7 (76.7–87.5)	[9]
Invertebrata: Crustacean	<i>Daphnia magna</i>	48 h	immobilization		>112	[9]
Invertebrata: Rotifera	<i>Brachionus calyciflorus</i>	24 h	mortality	130		[8]
Vertebrata: Fish	<i>Pimephales promelas</i>	96 h	mortality	31.8 (10.3–98.3)		[9]
PFHxA						
Algae	<i>Pseudokirchneriella subcapitata</i>	72 h	biomass	>100		[9]
Invertebrata: Rotifera	<i>Brachionus calyciflorus</i>	24 h	mortality	140		[8]
Invertebrata: Crustacea	<i>Daphnia magna</i>	48 h	mortality	>96.5		[9]
Vertebrata: Fish	<i>Oncorhynchus mykiss</i>	96 h	mortality	>99.2		[9]
Marine water						
PFBA						
Bacteria	<i>Photobacterium phosphoreum</i>	15 min	luminescence inhibition		14.07	[7]
PFPeA						
Bacteria	<i>Photobacterium phosphoreum</i>	15 min	luminescence inhibition		16.22	[7]
PFHxA						
Bacteria	<i>Vibrio fischeri</i>	30 min	luminescence inhibition		1335.39 ± 123.18	[58]
	<i>Photobacterium phosphoreum</i>	15 min	luminescence inhibition		17.20	[7]
Cyanobacteria	<i>Geitlerinema amphibium</i>	72 h	biomass <sup>f</sup>		995.49 ± 50.09 <sup>c</sup>	[45]
Algae	<i>Chlorella vulgaris</i>	72 h	biomass <sup>f</sup>		4019.51 ± 200.35 <sup>c</sup>	[45]
	<i>Skeletonema marinoi</i>	72 h	biomass <sup>f</sup>		1477.58 ± 72.00 <sup>c</sup>	[45]

Notes: hpf: hours post fertilization.

<sup>a</sup> test without pH adjustment.<sup>b</sup> test with pH adjustment.<sup>c</sup> standard deviation.<sup>d</sup> values based on combined sublethal and lethal embryotoxicity effect data.<sup>e</sup> malformation of head and tail and effects on growth.<sup>f</sup> biomass assessed by optical density.

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**Table 3**

PFBS acute aquatic toxicity (The full dataset is available in Table SM7, with references in SM10).

	Species	Exp time	Endpoint	LC50 (95% CI) mg L <sup>-1</sup>	EC50 (95% CI) mg L <sup>-1</sup>	References
Freshwater						
Sewage microorganisms		3 h	respiration inhibition		>1000	[10]
Cyanobacteria	<i>Anabaena</i> CPB4337	30 min	bioluminescence inhibition		8386	[48]
Algae	<i>Pseudokirchneriella subcapitata</i>	72 h	growth inhibition (biomass)		(7752–8693) >20250 (37% growth inhibition at 20,250 mg L <sup>-1</sup> )	[48]
	<i>Pseudokirchneriella subcapitata</i>	96 h	growth inhibition		5733	[10]
			biomass		(5659–5817) 2347 (2018–2707)	
Invertebrata: Crustacea	<i>Daphnia magna</i>	48 h	immobilization		2183 (1707–3767)	[10]
Vertebrata: Fish	<i>Danio rerio</i>	144 hpf	mortality and malformation effects <sup>a</sup>	1500 (1100–1900)		[56]
	<i>Danio rerio</i>	96 hpf	mortality and malformation effects <sup>a</sup>	>3000	450 (350–600) <sup>b</sup> 1900.78	[57]
		120 hpf		>3000	(1728.76–2089.92) <sup>b</sup> 1592.32 (1316.19–1776.96) <sup>b</sup>	
	<i>Pimephales promelas</i> (fathead minnow) <sup>c</sup>	96 h	mortality	1938		[10]
	<i>Lepomis macrochirus</i> (bluegill sunfish) <sup>d</sup>	96 h	mortality	(888–3341) 6452		[10]
Marine water						
Bacteria	<i>Vibrio fischeri</i>	15 min	bioluminescence inhibition		17,520	[48]
Invertebrata: Crustacea	<i>Mysidopsis bahia</i> (mysid shrimp)	96 h	mortality and abnormal behaviour		(16,850–18,200) 372	[10]
					(314–440)	

Notes: hpf: hours post fertilization.

<sup>a</sup> malformation of head and tail and effects on growth.<sup>b</sup> values based on combined sublethal and lethal embryotoxicity effect data.<sup>c</sup> juveniles 35 ± 5 mm and mean wet weight 320 ± 100 mg.<sup>d</sup> juveniles 44 ± 10 mm and mean wet weight 1000 ± 600 mg.

median of the HC5 is 22.2 mg L<sup>-1</sup>. To take into account the residual uncertainty, a default assessment factor of 10 is applied and an MAC-QS for the freshwater environment of 2.22 mg L<sup>-1</sup> is derived (Table 5).

Short-term L(E)C50 are available for marine algae, crustacea and fish and only one additional short-term toxicity data from a specific saltwater taxonomic group (Echinodermata) is available. In this situation HC5 calculated from the combined dataset is used to derive the QS<sub>sw,eco</sub> applying an additional AF of 5 was used, giving an MAC-QS for the marine environment of 0.450 mg L<sup>-1</sup> (Table 5).

### 3.2.2. MAC-QS<sub>water,eco</sub> for SC-PFAA

Short-term toxicity data are available for five taxonomic groups (bacteria, algae, plant, crustaceans, rotifers and fish) for PFBA, PFPeA and PFHxA and only for four taxonomic groups (bacteria, algae, crustaceans and fish) for PFBS. Though the acute base set (algae, Daphnia, fish) is complete, the range of taxonomic groups covered is insufficient to enable the use of SSD to derive the MAC-QS. Therefore, the assessment factor approach has been used for all the SC-PFAAs.

The TGD-EQS [1] recommend that, when there are at least 3 short-term tests of species from three trophic levels of the base set (algae, Daphnia, fish) and the standard deviation (SD) of the log transformed acute toxicity data is <0.5, the MAC-QS<sub>fw,eco</sub> is derived by applying an assessment factor of 10 to the lowest L(E)C50. Conversely, when the log transformed acute toxicity data SD is >0.5 the

MAC-QS<sub>fw,eco</sub> is derived by applying an assessment factor of 100 to the lowest L(E)C50. According to TGD-EQS, if no data for an additional marine taxonomic group are available, combined toxicity data are used to derive the MAC-QS<sub>sw,eco</sub> by applying an additional AF of 10 to the MAC-QS<sub>fw,eco</sub>.

The lowest acute toxicity concentrations for PFBA, PFPeA and PFHxA were obtained for the bioluminescence inhibition of the *Photobacterium phosphoreum* [7]. However this study was considered not reliable and excluded because its documentation is lacking. Furthermore there is no international agreement upon these methods and, on the consequence, these data would not be acceptable for use in ECHA dossiers.

From the reliable dataset for PFBA, the study on the freshwater invertebrate *Brachionus calyciflorus* (24 h LC50: 110 mg L<sup>-1</sup>) [8] has been chosen as the key driving study for MAC-QS setting. The SD of the log transformed acute toxicity data is >0.5. By applying an AF of 100 to the lowest acute effect concentration of 110 mg L<sup>-1</sup> an MAC-QS for the freshwater environment of 1.1 mg L<sup>-1</sup> is derived (Table 5). Because no data for an additional marine taxonomic group are available, an MAC-QS<sub>sw,eco</sub> for the marine environment of 0.110 mg L<sup>-1</sup> is derived (Table 5).

For PFPeA the SD of the log transformed acute toxicity data is <0.5. The study on the freshwater fish *Pimephales promelas* [9] is considered reliable without restriction and the 96 h LC50 (31.8 mg L<sup>-1</sup>) is used to derive the MAC-QS by applying an AF of 10, obtaining a MAC-QS for the freshwater environment of 3.18 mg L<sup>-1</sup>

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Table 4

PFOA and PFBS chronic aquatic toxicity (The full dataset is available in Table SM8, with references in SM10).

	Species	Exp time	Endpoint	EC10 (95% CI) mg L <sup>-1</sup>	NOEC mg L <sup>-1</sup>	References
Freshwater						
PFOA						
Algae	<i>Pseudochirchneriella subcapitata</i>	72 h	growth rate and biomass		200	[47]
		96 h			12.5	
	<i>Pseudochirchneriella subcapitata</i>	72 h	growth rate and biomass		200	[11]
		96 h			12.5	
Aquatic Plant	<i>Pseudochirchneriella subcapitata</i>	4.5 h	photosynthesis		413.06	[49]
	<i>Myriophyllum spicatum</i>	35 d	growth (plant length)	31.5 (0–68.2)	23.9	[60]
			root number	10.2 (6.6–13.7)	23.9	
			root length	8.8 (5.9–11.7)	23.9	
			longest root	24.3 (0–56.7)	23.9	
			node number	8.3 (5.3–11.4)	23.9	
			biomass (wet mass)	22.8 (0–53.5)	74.1	
			biomass (dry mass)	19.7 (0–53.5)	74.1	
	<i>Myriophyllum sibiricum</i>	35 d	growth (plant length)	23.7 (5.3–42.1)	23.9	[60]
			root number	29.2 (0–69.2)	23.9	
			root length	24.8 (0–64.1)	23.9	
			longest root	30.0 (0–90.6)	23.9	
			node number	7.8 (6.0–9.7)	23.9	
			biomass (wet mass)	21.6 (0–59.07)	23.9	
			biomass (dry mass)	24.7 (0–340.7)	23.9	
Invertebrata: Crustacea	<i>Daphnia magna</i>	21 d	reproduction		NR	[47]
			growth (as length)		44.2	
	<i>Daphnia magna</i>	21 d	reproduction		12.5	[52]
	<i>Moina macrocopa</i>	7 d	reproduction		3.125	[52]
	<i>Daphnia magna</i>	21 d	survival		>100	[61]
			reproduction		10	
	<i>Daphnia magna</i>	21 d	reproduction		20	[11]
			growth (as length)		44.2	
	<i>Daphnia magna</i>	21 d	reproduction rate		22	[11]
		14 d	reproduction		8	
			survival		60	
	<i>Daphnia magna</i>	21 d	survival	11.12		[50]
			reproduction	7.02		
Vertebrata: Amphibia	<i>Bufo gargarizans</i>	30 d	survival	5.89		[50]
Vertebrata: Fish	<i>Oncorhynchus mykiss</i>	85 d	mortality		40 <sup>c</sup>	[49]
	(rainbow trout) <sup>a,b</sup>					
	<i>Oncorhynchus mykiss</i>	85 d	mortality and growth (length)		40 <sup>c</sup>	[11]
	(rainbow trout) <sup>a,b</sup>					
	<i>Pimephales promelas</i>	39 d	survival		>74.1 <sup>d</sup>	[11]
			time for first oviposition		50 <sup>d</sup>	
			male plasma 11-ketotestosterone and testosterone		0.3 <sup>d</sup>	
	<i>Pimephales promelas</i> <sup>e</sup>	30 d	hatchability, survival growth and histopathology		>100	[11]
	<i>Pseudorasbora parva</i>	30 d	survival	11.78		[50]
PFBS						
Algae	<i>Pseudochirchneriella subcapitata</i>	96 h	growth inhibition		1077	[10]
			biomass		1077	
Invertebrata: Crustacea	<i>Daphnia magna</i>	21 d	reproduction/length		502	[10]
Marine water						
PFOA						
Algae	<i>Isochrysis galbana</i>	72 h	growth inhibition	41.6 (25.9–66.6)	25	[59]

## Notes:

<sup>a</sup> embryos.<sup>b</sup> larvae and juveniles.<sup>c</sup> for all life stages.<sup>d</sup> real concentrations applied to mesocosm.<sup>e</sup> eggs and fry.

(Table 5). Because no data for an additional marine taxonomic group are available, a MAC-QS<sub>sw,eco</sub> for the marine environment of 0.318 mg L<sup>-1</sup> is derived (Table 5).

The lowest acute toxicity concentration is 140 mg L<sup>-1</sup> (24 h LC50, *Brachionus calyciflorus*, [8]). The toxicity studies reported by [9] for PFHxA were not sufficiently documented and the toxicity values, which are maximum effect concentrations, cannot be used for QS derivation. Because three major taxonomic groups –Cyanobacteria, algae, and Rotifera–, but only two trophic levels are available, it is not possible to derive a consistent quality standard.

The lowest acute toxicity value for PFBS (96 h EC50 = 372 mg L<sup>-1</sup>) has been obtained for the crustacean *Mysidopsis bahia* [10] and the study has been considered reliable without restriction. The SD of

the log transformed acute toxicity data is >0.5. By applying an AF of 100 to the lowest acute effect concentration an MAC-QS for the freshwater environment of 3.72 mg L<sup>-1</sup> is derived (Table 5). *Mysidopsis bahia* is the only marine species represented in the data-set, since the *Vibrio* test is regarded as unreliable. In this situation the combined toxicity data can be used to derive the QS<sub>sw,eco</sub> and an additional factor of 10 is applied for saltwater, giving an MAC-QS for the marine environment of 0.372 mg L<sup>-1</sup> (Table 5).

3.2.3. AA-QS<sub>water,eco</sub> for PFOA

Long term toxicological data are available for five taxonomic groups including algae, plant, crustaceans, fish and amphibians together with a mesocosm study [11] (Table 4). No toxicological